

Genetic Fact Sheet

→ **Megrim**

Lepidorhombus whiffiagonis



2022





PANDORA

Paradigm for Novel Dynamic
Oceanic Resource Assessments

Gant agreement No: **773713**

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Genetic Fact Sheets

Review of available genetic information
on population structuring in exploited species

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PANDORA Project

The Blue Growth of European fisheries is at risk due to over-exploitation, unforeseen changes in stock productivity, loss of markets for capture fisheries due to aquaculture, future trade agreements opening European markets to external fleets, and fluctuations in the price of oil and other business costs. All of these risks need to be considered when providing advice needed to sustainably maximize profits for the diverse array of fisheries operating in European waters and to help safeguard the benefits this sector provides to the social coherence of local, coastal communities.

PANDORA aims to:

1. Create more realistic assessments and projections of changes in fisheries resources (30 stocks) by utilising new biological knowledge (spatial patterns, environmental drivers, food-web interactions and density-dependence) including, for the first time, proprietary data sampled by pelagic fishers.

2. Advise on how to secure long-term sustainability of EU fish stocks (maximum sustainable/"pretty good" and economic yields) and elucidate tradeoffs between profitability and number of jobs in their (mixed demersal, mixed pelagic and single species) fisheries fleets. Provide recommendations on how to stabilize the long-term profitability of European fisheries.

3. Develop a public, internet-based resource tool box (PANDORAs Box of Tools), including assessment modelling and stock projections code, economic models, and region- and species-specific decision support tools; increase ownership and contribution opportunities of the industry to the fish stock assessment process through involvement in data sampling and training in data collection, processing and ecosystem-based fisheries management.

The project will create new knowledge (via industry-led collection, laboratory and field work, and theoretical simulations), new collaborative networks (industry, scientists and advisory bodies) and new mechanisms (training courses and management tools) to ensure relevance, utility and impact.

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List of abbreviations

GFCM	General Fisheries Commission for the Mediterranean
ICES	International, Council for the Exploration of the Sea
IUCN	International Union for Conservation of Nature
IUU	Illegal, Unreported and Unregulated
MSY	Maximum sustainable yield
SAC	Scientific Advisory Committee
STECF	Scientific, Technical and Economic Committee for Fisheries
SD	Subdivision
TAC	Total Allowable Catch

North-East Atlantic ICES subareas, divisions and subdivisions

- 1) Subarea 1 – Barents Sea
- 2) Subarea 2 – Norwegian Sea, Spitzbergen and Bear Island
- 3) Subarea 3
 - Division 3.a, Skagerrak (subdivision 20) and Kattegat (subdivision 21)
 - Division 3.b-c, Sound (subdivision 23) and Belt Sea (subdivision 22)
 - Division 3.d, Baltic Sea (subdivisions 24-32)
- 4) Subarea 4 – North Sea (divisions 4.a-c)
- 5) Subarea 5 – Iceland (division 5.a) and Faroes Grounds (division 5.b)
- 6) Subarea 6 – West of Scotland (division 6.a) and Rockall (division 6.b)
- 7) Subarea 7
 - Irish Sea (division 7.a), West of Ireland (division 7.b), Porcupine Bank (division 7.c)
 - Eastern English Channel (division 7.d), Western English Channel (division 7.e)
 - Bristol Channel (division 7.f), Celtic Sea (divisions 7.g-h), Southwest of Ireland (divisions 7.i-k)
- 8) Subarea 8
 - North and Central Bay of Biscay (divisions 8.a-b)
 - South Bay of Biscay (division 8.c)
 - Offshore Bay of Biscay (division 8.d), West of Bay of Biscay (division 8.e)
- 9) Subarea 9 (Portuguese Waters)
- 10) Subarea 10
 - Azores Grounds (division 10.a) and Northeast Atlantic South (division 10.b)
- 11) Subarea 11 (incorporated in FAO Fishing Area 34)
- 12) Subarea 12 North of Azores
 - southern mid-Atlantic Ridge (division 12.a)
- 13) Subarea 13 (incorporated in FAO Fishing Area 34)
- 14) Subarea 14 East Greenland, Northeast Greenland (14.a), Southeast Greenland (14.b)

How to read the factsheets

Genetic structure factsheets are presented for each species. Current knowledge on genetic population structure is summarised and compared with stock units used in assessment and management. The presence of mismatches is emphasised as well as priorities for future work. At the beginning of the factsheets, a summary is presented with green-yellow-red color symbols for 'Population structure', 'Match between genetic and stock assessment units' (units for which scientific advisory bodies, as ICES and the GFCM, provide advice on stock status and fishing opportunities), 'Match between genetic and management units' (units for which TACs are set by the European Council), 'Match between stock assessment and management units'. The information in the factsheet is organized in the following sections:

Distribution: general information can be found on the distributional range of the species, with a focus on the NE Atlantic, Mediterranean and Black Sea.

Current management status: an overview is provided on the current management and assessment units present for the species in European Seas. The importance of the species for each fishery is included, reporting if the species is mainly a by-catch or if direct fishery exists for the stocks. A mismatch between stock assessment and management units already exists for certain species and it is showed in Table 2.

Genetic population structure in a nutshell: provides the key take-home messages, both in terms of current knowledge on genetic population structure and in terms of priorities for future work. In this section, an overall picture of population structure of the species is given, based on considerations on the type of markers, sampling designs and findings of the included studies. It is also discussed if genetic evidence supports the stock assessment and management units currently in use.

Mismatch: in this section the mismatch between genetic and stock assessment/management units is highlighted. Two types of mismatch can be observed. Here, we refer to 'Type I' mismatch when a genetically homogeneous population is assessed/managed in multiple stock units (oversplitting); while we refer to 'Type II' mismatch when genetically different populations are wrongly considered part of the same stock assessment/management unit (undersplitting).

Summary of genetic evidence: in this section a more detailed summary of the studies is provided in a chronological way. In general, the type of genetic markers used by different studies depends on the widely available markers at the time. Early studies used allozymes and often reported a lack of differentiation among sample locations. However, later studies using the more highly polymorphic microsatellites and SNPs showed presence of differentiation even in areas where it was not previously detected. Conversely, in other cases presence of differentiation was reported at few allozyme loci, not confirmed

subsequently with strictly neutral markers. This and other contradictions between studies were addressed if possible. Advances in sequencing technology, as well as the use of more sophisticated statistical analysis and sampling design to maximise the detection of population structure have made enormous changes in the awareness we have of genetic structure in marine fish species (Hauser and Carvalho, 2008). Most of the mismatches found in initial studies between genetic population structure and stock assessment and management units were due to a lack of differentiation reported between samples assessed/ managed in different units (referred to as 'Type I' mismatch in Table 1). However, these mismatches are often solved by more recent investigations, that applied highly polymorphic markers, as well as a sampling design that maximise the chance of detecting population structure, i.e. collecting individuals in spawning aggregations. Particular emphasis should be placed on the sampling season and individuals included in the analysis that are extremely important factors for the detection of population structure in marine fish species (Nielsen et al. 2009b). Moreover, despite in previous studies a neutral background of low differentiation was commonly detected, recently the application of markers under selection allowed the detection of high levels of differentiation and occurrence of locally adapted populations. Therefore, a summary of genetic studies found in literature is provided. For each study, sampling design, temporal and spatial analyses and markers used have been critically evaluated. Strengths and shortcomings of the available studies are reported and based on these considerations an overview is given.

Table 2.1. Summary table of available information on genetic population structure and match between genetic, assessment and management units of commercial fish species exploited in the NE Atlantic, Mediterranean and Black Sea.

Species	No. Studies	Population structure	Match genetic-Stock assessment units	Match genetic-management units	Match stock assessment-management units	IUCN status
Four-spot megrim, <i>Lepidorhombus boscii</i>	2	yes	Yes	no	no	LC

IUCN Abbreviations: NE= Not evaluated, DD= Data Deficient, LC= Least Concern, NT= Near Threatened, VU= Vulnerable, EN= Endangered, CR= Critically Endangered. Eu= Europe, Glo= Global, Med= Mediterranean (IUCN 2021).

FACT SHEET

Megrim, *Lepidorhombus whiffiagonis*

Number of studies	3
Population structure	✓
Match genetic- Stock assessment units	✗
Match genetic- Management units	✗
Match Stock assessment- Management units	✗



Distribution¹

Megrim, *Lepidorhombus whiffiagonis* (Walbaum, 1792), is a demersal flatfish species widely distributed in the North-East Atlantic, from Icelandic and Faroese waters to Cape Bojador, as well as in the Mediterranean Sea (Garcia-Vazquez et al. 2006). It is caught, together with four-spot megrim in mixed fisheries directed to other demersal species, mainly hake, anglerfish and *Nephrops*.

Current management status

Megrim is mainly a by-catch species in demersal fisheries directed to whitefish and flatfish. Two stocks of megrim are identified by ICES: one in the Celtic Seas and part of Bay of Biscay (8.a-b, d) and the other in southern Bay of Biscay and Atlantic Iberian waters (Figure 3.5). Combined TACs exist for megrim and four-spot megrim for combined TACs), and ICES recommended the set of separate TACs to avoid overexploitation and secure a more sustainable fisheries management. Both species are part of the EU multiannual plan (MAP) for Western Waters and adjacent waters (ICES, 2020e; and references therein). Landings for megrim in 2019 were 239 t for the Iberian stock (ICES

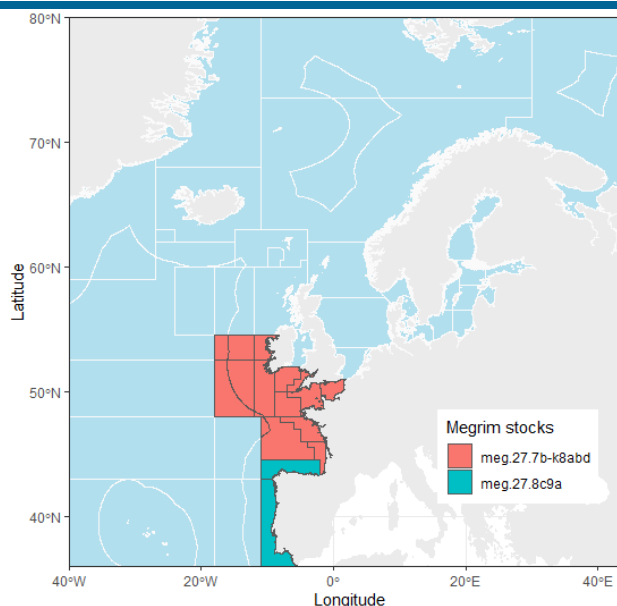


Figure 3.5. Megrim ICES stock assessment units

¹ Further details on symbols and how to read the factsheet are provided on page 16



2020j), while in the Celtic Seas and Bay of Biscay preliminary landings were 12164 t (ICES 2020i).

Genetic population structure in a nutshell

In line with assessment and management units, the differentiation between the northern (division 6) and southern NE Atlantic (8.c and 9) is supported by genetic analysis, as well as the divergence between northern and southern Bay of Biscay. However, further studies are needed to understand patterns of population structure and spatial distribution of megrim in the Bay of Biscay and Celtic Sea. Genetic evidence supports the presence of different stock assessment units of megrim in the Rockall (division 6.b) and in the northern North Sea, west of Scotland (division 4.a and 6.a). Moreover, substructure was suggested in the northern North Sea and West of Scotland, currently considered part of the same stock by ICES.

Mismatch

Mismatch between stock assessment and genetic units is present for the Bay of Biscay and Celtic Sea that indicated genetically distinct units, currently part of the same stock assessment unit. Further studies are needed to elucidate genetic structure in these divisions, since the Bay of Biscay appeared more similar to northern divisions of subarea 6 while the Celtic Sea exhibits closer similarity to the southern Iberian stock.

Genetic differences were found at microsatellite loci (Macdonald & Prieto) between the northern North Sea and West of Scotland, currently considered part of the same stock by ICES. If additional analyses confirm this difference, appropriate stock units reflecting the biological populations present are recommended. Moreover, from a fisheries management perspective, a TAC is given for the Rockall and west of Scotland jointly despite that genetic differentiation of Rockall megrim was showed, resulting in a mismatch.

Summary of genetic evidence

A total of three studies investigating genetic population structure of megrim are present in the literature (Table 1). Garcia-Vazquez et al. (2006) using a combination of nuclear and mitochondrial markers detected genetic differentiation between megrim inhabiting the NE Atlantic and the Mediterranean Sea with the strait of Gibraltar acting as potential barrier to gene flow. At a finer scale, Danancher and Garcia-Vazquez (2009) developed a set of highly polymorphic markers that not only supported the differentiation between the NE Atlantic and the Mediterranean, but also indicated the presence of structure within the NE Atlantic (global F_{ST} 0.158, $P < 0.001$). Microsatellites supported the presence of at least two populations, a northern, in division 6, and a southern including south Bay of Biscay and Portuguese waters (division 8.c and 9). Difficulties arose for samples from areas between these two stocks. Megrim from division 7 clustered with the southern



stock (Division 8.c and 9) while samples from divisions 8.a-b, d clustered with the northern stock (Subarea 6). However, the sampling season is not reported and if sampling occurred outside the spawning season, there is a reduced chance of identifying genetic structuring. Further studies are needed to better understand patterns of population structure and spatial distribution of megrim in these areas.

Furthermore, population structure of megrim in the Northern shelf was evaluated in a report from the NAFC Marine Centre (Macdonald & Prieto). Currently, ICES recognize two stocks of megrims (*Lepidorhombus* spp.): one in the northern North Sea and west of Scotland (Division 4.a and 6.a) and the other in the Rockall (Division 6.b). The analysis through microsatellites supported the presence of these stocks and moreover detected further localised differentiation between west of Scotland (6.a) and the northern North Sea (4.a) with mixing in the northeastern part of division 6.a and the eastern part of division 4.a. Further studies are needed to confirm the differences between northern North Sea and West of Scotland and the relative patterns of mixing.

Table 1. Summary table of genetic population structure studies of commercial marine fish species exploited in the North-East Atlantic Ocean, Mediterranean and Black Sea.

Species	Region	Sampling locations	No. Samples (Number of individuals)	Spawning	Maturity	Life stage	Genetic Marker	Differentiated	Mismatch genetic-SA	Mismatch genetic-MZ	LA, LG, MSA	Reference
Megrim	NE Atlantic, Med	w SCO (1), EC (1), BOB (2), PRT (1), Med (1)	6 (303)	na	na	Ad	mtDNA (16S); nDNA (5S)	Yes	Type I	Type I		(Garcia-Vazquez et al. 2006)
	NE Atlantic, Med	w SCO (1), CS (2), BOB (2), PRT (1), Med (1)	7 (191)	na	na	Ad	Msat (6)	Yes	Type II	Type II	LG	(Danancher & Garcia-Vazquez 2009)
	NE Atlantic	Roc (3), NS (3), w SCO (3)	9 (270)	y	na	Ad	Msat (6); mtDNA (1)	Yes	Type II	Type II	LG	(Macdonald & Prieto)

The following abbreviations are used for the geographic locations: North-East Atlantic (NE Atlantic), Mediterranean Sea (Med), Northwest Atlantic (NWA), Adriatic Sea (Adr), Aegean Sea (Aeg), Africa (AFR), Alboran Sea (Alb), Atlantic (Atl), Atlantic Iberian (Atl IB), Australia (AU), Azores (Azo), Baltic Sea (BAL), Barents Sea (BS), Bay of Biscay (BOB), Black Sea (BLS), British Isles (BI), Canada (CAN), Canary (Cn), Cantabrian Sea (Cant), Celtic Sea (CS), English Channel (EC), Faraday Seamount (Far), Faroe Islands (FRO), fjord (fj), Galicia (Gal), Greece (GRC), Greenland (GRL), Gulf of Cadiz (GC), Gulf of Lion (GoL), Hebrides (Heb), Iceland (ICE), Ionian Sea (Ion), Ireland (IRE), Irish Sea (IS), Irminger Sea (Irm), Kattegat (Kat), Lake Mogilnoe (Mog), Lofoten (Lof), Madeira (Mad), Marmara Sea (MS), Mid-Atlantic Ridge (MAR), Morocco (MOR), Namibia (Nam), New Zealand (NZL), Newfoundland and Labrador (NL), North Sea (NS), North Sea-Baltic Sea Transition zone (NBTZ), Norway (NOR), Nova Scotia (Nov), Porcupine Bank (Por), Portugal (PRT), Reykjanes Ridge (Reyk), Rockall Bank (Roc), Russia (RUS), Scotian Shelf (SS), Scotland (SCO),



Shetland (SHE), Sicily (SIC), Skagerrak (Ska), Spain (SPA), Svalbard and Jan Mayen (SJM), Tasman Sea (TS), Tunisia (TUN), Tyrrhenian Sea (Tyr), White Sea (WS); north (n), south (s), east (e), west (w), central (c); Norwegian Coastal Cod (NCC), North-East Arctic Cod (NEAC).

Sampling locations (for abbreviations see below) and in brackets the number of samples are shown; the total number of samples and individuals analysed is reported, as well as the number of temporal replicates in superscript or (*) if multiple temporal replicates are included. The spawning, maturity and life-stage of samples included are summarised as follow, *Spawning*: y= if samples collected in spawning season/grounds are included, na= not available, no= samples outside spawning season/grounds. *Maturity*: y= mature individuals included; na= maturity not available; no= immature individuals. *Life-stage*: Ad= adult; juv= juveniles; lar= larvae; eg= eggs; na= not available. Genetic markers (All= allozymes; Msat= microsatellites; Minisat= minisatellites; SNPs= Single Nucleotide Polymorphisms; mtDNA= mitochondrial DNA; Cyt-b= cytochrome b; COI= Cytochrome c Oxidase subunit I; COIII= Cytochrome c Oxidase subunit III; CR= Control Region; RAPD= Random Amplified Polymorphic DNA); number of loci or base pairs analysed in brackets, in superscript S= if at least one locus is under selection, N= neutral markers (only if neutrality was tested). Differentiation, if genetic differentiation was detected (Yes, No). Mismatch genetic- SA= mismatch of the genetic units found and the stock assessment units. Mismatch genetic- MU = mismatch of genetic units with the management units. We refer to 'Type I' mismatch when a genetically homogeneous population is assessed/managed in multiple stock units (oversplitting); while we refer to 'Type II' mismatch when genetically different populations are wrongly considered part of the same stock assessment/management unit (undersplitting). LA= Local Adaptation, LG= Landscape Genetics, MSA= Mixed Stock Analysis.

Table 2. Mismatch between stock assessment (SA) units and genetic population structure (Type I and II explained) and mismatch between management and genetic units.

Species	Assessment unit	Mismatch SA unit - genetics (Type II)	Mismatch SA unit -genetics (Type I)	Management units	Mismatch management unit - genetics
Megrim spp., <i>Lepidorhombus spp.</i>	lez.27.4a6a	Differentiation between 4.a and 6.a, likely for megrim (Macdonald & Prieto)			
	lez.27.6b			- 5b ^{U,I} ; 6; 12, 14 ^I	Megrim in 6.a and 6.b, genetically different, managed in one TAC (Macdonald & Prieto)